



The role of Nanoparticles for Reactive Oxygen Species (ROS) in Biomedical Engineering

Sargol Mazraedoost¹, Reza Masoumzadeh², Zahra Javidi¹, Yousef Ashoori¹

¹ Biotechnology Research Center, Shiraz University of Medical Sciences, Shiraz, Iran.

² Department of Medical, Shiraz University of Medical Sciences, Shiraz, Iran.

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Abstract

Among the greatest varied and cross-cutting features of biomedical nanotechnology applications is the synthesis, design, and characterization of novel nanomaterials. New developments in synthetics and engineering make it possible to produce an extensive variety of nanoparticles (NPs) and biocompatible nanostructured materials widely used in efficient diagnosis, drug delivery, and therapeutic procedures or deprived of other chemical and/or surface modifications of biomolecules. Because of their physical and chemical properties, metal-based nanoparticles (MNPs), as well as quantum dots (QDs), magnetic NPs, metal NPs, and metal oxide NPs, have a tremendous amount of power for biomedical applications. Nanoparticles (NPs) have superior (chemical and physical) features that create an ideal for different usages. Metallic NPs' structural modifications result in various biological activities, leading to diverse development capacities for reactive oxygen species (ROS). With chemistry, size, surface area, and particle shape, the amount of ROS provided by metallic NPs are correlated. In cell biology, ROS has many functions. ROS generation is a critical component in the toxicity caused by metallic NP and cellular signaling in cell differentiation, proliferation, and death.

Keywords: *Metal nanoparticles; Nanomaterials; Reactive oxygen species; Nanoscience; ROS Generation and Oxidative Stress*

1 Introduction

Nanoscience discusses the study and use of small materials through measurements. It may take advantage of many other fields similar to or less than 100 nm, for instance, physics, material science, engineering, biology, medicine, and chemistry. Analysis and development of nanoparticles (NPs) and nanostructured materials (NSMs) [1] In this field is one of the most active research areas. Electrical and thermal conductivity, light absorption, catalytic activity, and dispersion are just a few of the unique physicochemical properties of NPs and NSMs that allow several different fields to benefit from improved performance, starting with bulk counterparts such as agriculture, food, medicine, and cosmetics [2-8]. Appropriate applications for NSMs and NPs in fluorescent biological products have been described in the latter field. Labeling [9, 10], tissue engineering [11], protein analysis [12], pathogen detection [13], DNA structure testing [14], purification and cell and molecular biological separation [15], MRI contrast reinforcement [16], gene and drug delivery system [17-19]. In particular, NPs have been used successfully in recent decades in the clinic as important different therapy methods, for instance, photothermal therapy (PPT) [20], High-Intensity Focused Ultrasound (HIFU) [21], sonodynamic therapy (SDT) [20, 22-25] and photodynamic therapy (PDT) [26-28].

These treatments are becoming more popular due to their capacity to cause prokaryotic and eukaryotic cells to die via fundamental biological processes such as the generation of NP-mediated ROS [29]. Producing ROS and reducing scavenging pathways once secretion into the body via various internalization techniques such as skin absorption, parenteral, inhalation administration, and oral some NPs may influence redox homeostasis [6, 19, 30-50].

2 The production of reactive oxygen species (ROS) and the mechanism of nanotoxicity

A primary mechanism contributing to nanotoxicity is subsequent production of oxidative stress and ROS formation, which includes promotion, cancer start, cytotoxicity, apoptosis, changes in cell mobility, not controlled cell signaling, and DNA damage [51-53]. The extent to which engineered nanomaterials produce ROS depends on the nanoparticles' chemical nature [54]. Engineered nanomaterials have a greater specific surface area, smaller size, and more surface reactivity than bulk-size equivalents, resulting in increased ROS levels and cytotoxicity [55]. In various biological systems, such as human erythrocytes and skin fibroblasts, a range of NMs has been reported to produce cytotoxicity mediated via ROS [56]. Winnik and Maysinger [57] QDs caused cell damage and oxidative stress, mediated with ROS. Akhtar et al. [58] Silica NPs have been used to cause subsequent oxidative stress and cytotoxicity in a dose-dependent procedure, intermediary via lipid peroxidation

Corresponding author: Sargol Mazraedoost, Biotechnology Research Center, Shiraz University of Medical Sciences, Shiraz, Iran. Sargol.mazraedoost7@gmail.com

and ROS production inside the cell coat. Akhtar et al. [59]. Also determined that nano-CuO induces releases lactate dehydrogenase (LDH), cellular toxicity in mouse fetal fibroblasts (BALB-3T3), ROS induction, and lipid peroxidation are the mechanisms that cause stress oxidative in a dose-dependent procedure. As further evidence of this mechanism, Nano-ZnO has been shown to persuade cellular toxicity, which is primarily intermediary via ROS induction, creating oxidative harm, the release of inflammation mediators, and cell decease inside phagocytosis of RAW 264.7 cell line, as well as the alteration in people broncho-epithelial cells (BEAS-2B) [52, 60]. Nano-TiO₂ also generates ROS, contributing to photocytotoxicity and cytotoxicity, close to nano-ZnO [61-65]. For biological purposes, both gold cobalt nanoalloys and AuNPs have been investigated. Girgis et al. [66] The increase in micronucleus formation 8-OHdG production was linked to Au-Co nanoalloy induced cancer primer gene changes in mice.

In contrast, these harmful actions caused by AuNPs were significantly reduced. Girgis et al. [66] claimed that increased oxidative stress is to blame for these destructive behaviors. Hsin et al. [67] revealed that AgNPs motivated apoptosis in NIH3T3 cells, intermediary with a mitochondrial pathway C-Jun-terminal-kinase-dependent and ROS mechanism. Mei et al. [6] In mouse lymphoma cells, AgNPs caused oxidative stress and mutation, intermediary via ROS production. Kim et al. [68] In animal tissues and cultured cells, AgNPs created apoptosis, oxidative stress, and genotoxicity. Shvedova et al. [69] High-dose, single-walled CNT incubation in bronchial epithelial cells and keratinocytes has been documented to produce ROS, oxidative stress, lipid peroxidation, and mitochondrial malfunction.

Superparamagnetic Fe₂O₃ NPs are cellular toxicity and cause apoptosis and ROS [70]. Wang et al. [71] Hepatocytes from catfish and human HepG2 cells, Cytotoxicity of four nano metallic oxides, nano-CuO, nano-Co₃O₄, nano-ZnO, and TiO₂. nano, was contrasted. The mediated toxicity in both cell systems was in CuO > ZnO > Co₃O₄ > TiO₂; the induced cellular toxicity is caused by damage to mitochondrial membranes and the cells' and ROS. The cytotoxicity of HepG2 cells was more significant than that of catfish primary hepatocytes. As displayed in (Figure 1), damage to the mitochondrial and cells membranes and ROS are responsible for the induced cellular toxicity. HepG2 cells had higher cytotoxicity than catfish primary hepatocytes. ROS does not mediate all the toxicity of nano metallic oxide. The research by Karlsson et al. is an excellent example. [72]. They concluded oxidative stress, cytotoxicity, and the DNA damage of diverse nano metal oxides (ZnO, TiO₂, CuO, Fe₂O₃, CuZnFe₂O₄, and Fe₃O₄), MWCNTs and carbon nanoparticles, in people epithelial's lung cells (A549) [73]. CuONPs were the most effective in motivating cellular toxicity, oxidative lesions, substantially raising intracellular ROS, and DNA damage; ZnONPs exhibited DNA damage and cytotoxicity. Nano-TiO₂NPs, which included both types of anatase and rutile, induced only DNA damage. NanoFe₃O₄ and Fe₂O₃NPs were shown to have nil or minimal cytotoxicity. NanoCuZn Fe₂O₄ induced DNA damage that was effective. CNTs were also shown to cause DNA damage and cytotoxicity. These outcomes indicate that CuONPs are the only nanomaterial investigated that generates ROS and has the highest cytotoxicity and genotoxicity[72].

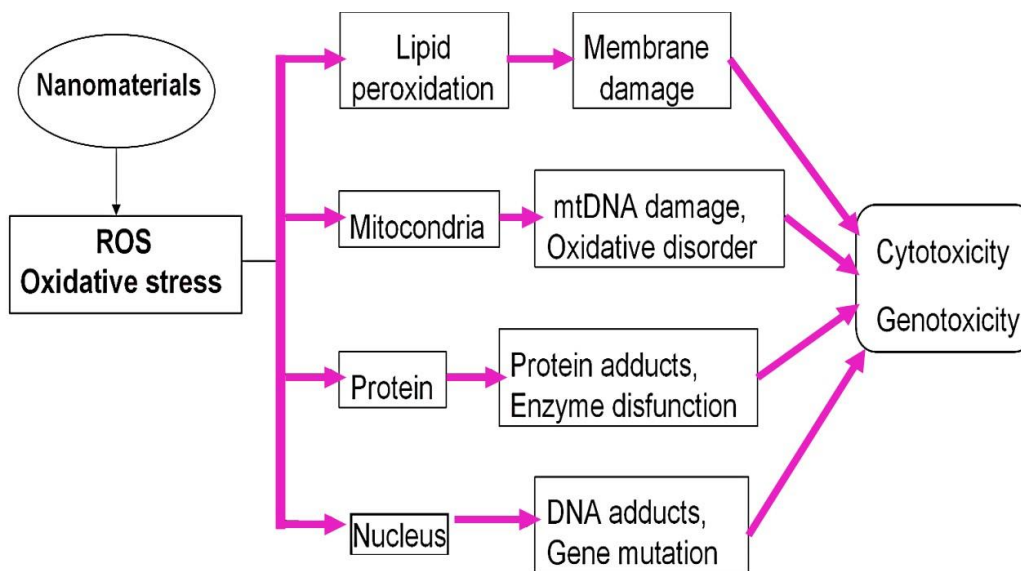


Figure 1. ROS generation-mediated nanomaterial-induced toxicity [73].

3 Oxidative Stress and the Production of Reactive Oxygen Species (ROS)

While homeostasis and cell signaling are ROS, the primary signaling molecules create oxidases in cells, resulting from the excitation and uniform decrease in molecular oxygen, hydrogen peroxide, hydroxyl radicals, and superoxide anion [74]. Molecular oxygen generates the primary ROS and superoxide anion by lowering one electron catalyzed with NADPH oxidase. Dismutation of oxygen and the metal-catalyzed Fenton reaction can produce hydrogen peroxide or hydroxyl radicals, respectively [75, 76]. Several of the endogenous ROS roots are peroxisomes, inflammatory response, mitochondrial respiration, and microsomes. Free radicals produced as a result of Fenton-type transition metal ion-catalyzed processes and important by-products of mitochondrial respiration, on the other hand, can essentially control numerous signal transduction pathways in a dose-dependent manner. Increased ROS levels cause nucleic acid and lipid peroxidation and oxidation due to necrosis and cellular death. Low or medium ROS levels, on the other hand, enhance mitogenic signaling via reversible oxidation [6, 77-80]. Together with non-free radical and free-radical oxygen-comprising molecules, there are sulfur species, copper (Cu), iron (Fe), and reactive nitrogen, which might be attributed to enhanced oxidative stress and ROS production, disrupting the redox equilibrium [81-83]. In this respect, it is the responsibility of antioxidant molecules, for instance, vitamin E, glutathione (GSH), flavonoids, ascorbic acid, and detoxifying enzymes, for example, glutathione peroxidase (GPX), Superoxide dismutase (SD), and catalase (CAT), and to manage the suitable physiological amount of ROS (SOD) [82]. According to this concept, Antioxidant enzyme systems respond to growing oxidative stress levels in cells and tissues. Transcription of phase II antioxidant enzymes (in the presence of moderate oxidative stress) is activated by induction of nuclear factor (derived from red blood cells 2) -like 2 (Nrf2). At a reasonable level, proinflammatory responses are produced by the nuclear factor kappa-light-chain activated B cell enhancer (NF- κ B) and redox-sensitive MAPK. High amounts of oxidative stress, on the other hand, cause damage to the electron chain malfunction and mitochondrial membrane, resulting in cell dying [74].

Consequently, disturbance of the usual redox condition leads to the development of peroxides and free radicals that have harmful impacts on cell parts such as lipids, DNA, and proteins [84], resulting in cell death, fibrosis, and carcinogenesis (Figure 2) [85-87].

ROS plays an essential role in cell signaling, differentiation, cell death, cell survival regulation, and factor development associated with inflammation [88, 89]. Biologically-significant ROS components such as free radicals, for example, hypochlorite (OCl^-), nitric oxide (NO), peroxomonocarbonate (HOOCO_2^-), peroxyxynitrous acid (ONOOH), peroxyxynitrate (O_2NOO^-), organic peroxides (ROOH), peroxyxynitrite (ONOO^-), hypochlorous acid (HOCl), ozone (O_3), hypobromous acid (HOBr), Superoxide ($\text{O}_2^{\cdot-}$), nonradicals, such as hydrogen peroxide (H_2O_2), carbon dioxide radical ($\text{CO}_2^{\cdot-}$), (RO^{\cdot}), peroxy (RO_2^{\cdot}), hydroperoxyl (HO_2^{\cdot}), carbonate ($\text{CO}_3^{\cdot-}$), hydroxyl (HO^{\cdot}), singlet oxygen ($^1\text{O}_2$), and alkoxy [29, 90, 91].

$\text{O}_2^{\cdot-}$ is a short-lived free radical due to its fast conversion to H_2O_2 , intermedia via SODs[92]. One non-radical ROS analogues are H_2O_2 , which has higher stability and longer biological life than free radicals [93]. Superoxide is produced by NADPH oxidase (NOX) and mitochondria, which may begin lipid peroxidation or deactivate certain enzymes (Figure 3A) [94]. Electrons produce superoxide due to incomplete reduction of O_2 , and finally, H_2O_2 is produced from SOD, which is an important defense against antioxidants in almost every cell's exposure to O_2 . Three different types of SOD are included SOD3 (a zinc and copper ion-comprising extracellular SOD), SOD2 (an Mn ion-comprising mitochondrial SOD), and SOD1 (a zinc and copper ion-comprising SOD mainly found in the cytoplasm). In different physiological conditions, ROS is generated as Intermediaries, and its cellular amounts are greatly adjusted with different detoxifying enzymes, including glutathione peroxidase (GPX), SOD, and catalase (CAT), or with different antioxidants, containing glutathione (GSH), ascorbic acids, flavonoids, and vitamin E [90]. There are strong links between ROS and cellular pathology, metabolism, and production. [95, 96].

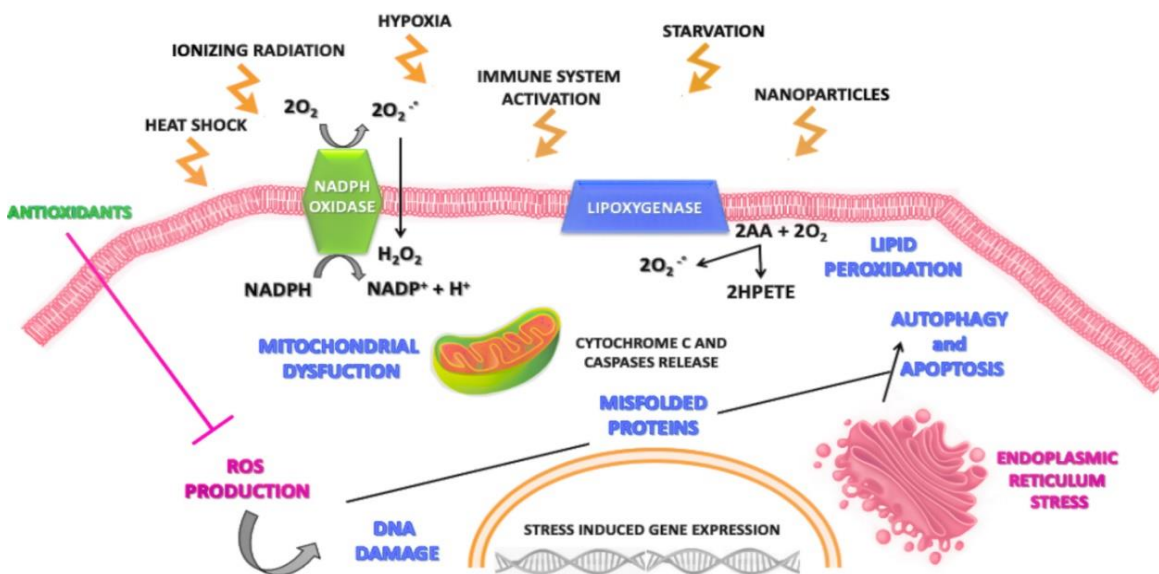


Figure 2. Different triggers for producing ROS-induced routes and ROS that lead to cell harm are depicted in this diagram [31].

The redox without balance is a flaw in the balance between ROS production and their neutralization by cellular antioxidant agents. Interference with oxidative damage or cell signaling to macromolecules, like lipids, nucleic acids, and proteins produces detrimental effects on cells when redox homeostasis is disrupted [97]. Modulating ROS production, on the other hand, enhances the stimulation of main signaling molecules that control survival, differentiation, proliferation, and cell death [98-101]. In nanotechnology, small materials (dimensions <100) are most used. [102]. In the 21st century, this field has grown rapidly, with steady development in new applications. Engineered nanoparticles (NPs) have unique physicochemical features produced for various biological and industrial functional applications [54]. The particular thermal, electrical, chemical, and biological properties of NPs make them useful in a wide range of manufacturing, cosmetics, medical, agricultural, clothing, and food industry applications [61, 103, 104]. NPs' physicochemical features' total diversity often provides investigation opportunities related to their toxic effects [105]. Via different routes and oral intake, inhalation, and skin absorption, the internalization of NPs into the human body can be promoted. After absorption, they enter the biological field, and body fluids are the site of communication between cellular components. Corona protein is caused by cellular molecules coating NP surfaces [8, 106, 107]; this enables the biological identity of NPs to be recognized [108]. Extensive studies on commercial NPs, optical toxicity and cytotoxicity of nanotechnology need to be developed to better nanotechnology. NPs have toxic properties that lead to oxidative stress, inflammation, inhibition of cell division, genetic damage, and cell death [109, 110]. Inflammation, fibrosis, genotoxicity, and carcinogenesis are all pathogenic processes that NP-mediated ROS triggers. For instance, the physicochemical properties of NPs, for instance, surface area, size, chemical structure, and charge, influence it [111]. Increased production of pro-inflammatory and fibrotic cytokines and stimulation of inflammatory cells like neutrophils and macrophages can induce NP-related toxicity, resulting in increased ROS production [112-114]. The mechanism associated with the generation of NP-induced ROS differs between the core cellular mechanism associated with ROS development and diverse NPs not specified. Most metal-based NPs via Fenton-type reactions may cause free-radical-facilitated toxicity [7, 19, 29, 115, 116].

4 Reactive Oxygen Species (ROS) Generation Sources

Mitochondria, microsomes, peroxisomes, NOX complexes, and endoplasmic reticulum (ER) (7 separate isoforms) in cell coat are the primary sources of intracellular ROS (Figure 3A) [76, 117]. Mitochondria are the essential intrinsic source of ROS production through electron transport's mitochondrial mechanism (Figure 3B) [118]. Increased cytoplasmic calcium accumulation results in ROS development and mitochondrial electron transport chain. Throughout the mitochondrial processing of water and ATP, minor amounts of O₂ are released after ROS development's early stages. The activity of Complex III (cytochrome/ubiquinone c reductase, bc₁ complex, and coenzyme Q) and Complex I (NADH ubiquinone oxidoreductase) The first component of ROS made by mitochondria is the superoxide anion found in the mitochondrial membrane and matrix [119, 120]. Metals in the gap between the membranes include Zn-SOD, Cu, and Mn, which catalyze a

stable form of H₂O₂ from superoxide anions [121]. Other possible causes of mitochondrial ROS production include monoamine oxidase and alpha-ketoglutarate dehydrogenase [122, 123]. NOX is a non-mitochondrial generator of ROS and Through oxygen reduction, which is interceded with the electron donor NADPH, plays a critical role in superoxide production. Mammalian NOX consists of seven isoforms DUOX₂, DUOX₁, and NOX1-5, most of which produce superoxide, while DUOX₂, DUOX₁, and NOX₄ produce H₂O₂ [124, 125]. One of the cellular organs that play a role in the production of ROS is ER. The ideal oxidizing conditions are provided in the ER lumen to provide disulfide bonds and protein accumulation [126]. The ROS production process involves several biological enzymes containing NO synthase, lipoxygenase, xanthine oxidoreductase, cyclooxygenase, and cytochrome P450 monooxygenase. ONOO⁻'s a nitrating agent and potent oxidizing, outcomes from the interaction amongst O₂⁻ and NO [127]. ROS-inducing agents, for example, toxins, radiation, and exposure to nanomaterials, are extracellular causes of ROS generation (Figure 3A) [75]. Against microbial invasion, disease, and pollutant toxicity, oxidative stress activates defensive mechanisms in macrophages and neutrophils. Given the Fenton reaction's iron, which generates hydroxyl radicals, Fe²⁺ (free iron) is a crucial component connected to ROS generation's toxicity [128, 129].

5 ROS Generation Induced by NPs and the Mechanisms Involved

ROS generation related to NP is controlled with the variables described: physical properties (surface area and size), particle chemistry, and internalization of NPs [72, 130]. This section describes the mechanism of ROS related to NP generation, including the role of NP-related items in ROS the interaction and creation of NPs with cellular ingredients (Figure 3B).

6 Important Roles of NPs in ROS Production

Previous studies have extensively described ROS production's toxic and hazardous effects by different nanomaterials with different chemical structures. [75, 131]. NPs have complete physicochemical specifications (aggregation status, solubility, shape, size, and surface area) related to their bulk of origin or microparticles, linked to their ability to generate ROS [51, 52, 132-135]. NPs vary substantially from their mass counterparts in surface area, significantly more prominent and typically containing a better percentage of atoms [135]. Particle weight and particle size, representing the volume/surface ratio, are reversely related [135]. The amount of reactive points on the NP's surface is restricted by particle size since smaller NPs have higher surface areas and mass [136-138]. Large NPs dramatically accelerate chemical reactions. The dangling bonds of atoms on the NP surface enhance NP-induced biological catalysis, resulting in high chemical reactivity (immobilized free radicals) [139]. Compared to more significant NPs, minor NPs contribute to constructive changes and improvements in the electrical characteristics of the particle level, finally leading to reactive groups on particle surfaces [140, 141]. Due to differences in surface characteristics, previous investigations have revealed that ZnO NPs and silicon NPs of similar shape and size exhibit various toxicity levels. ZnO NPs, for example, have more chemical activity than SiO₂-NPs and, as a result, create better amounts of oxidative stress produced with

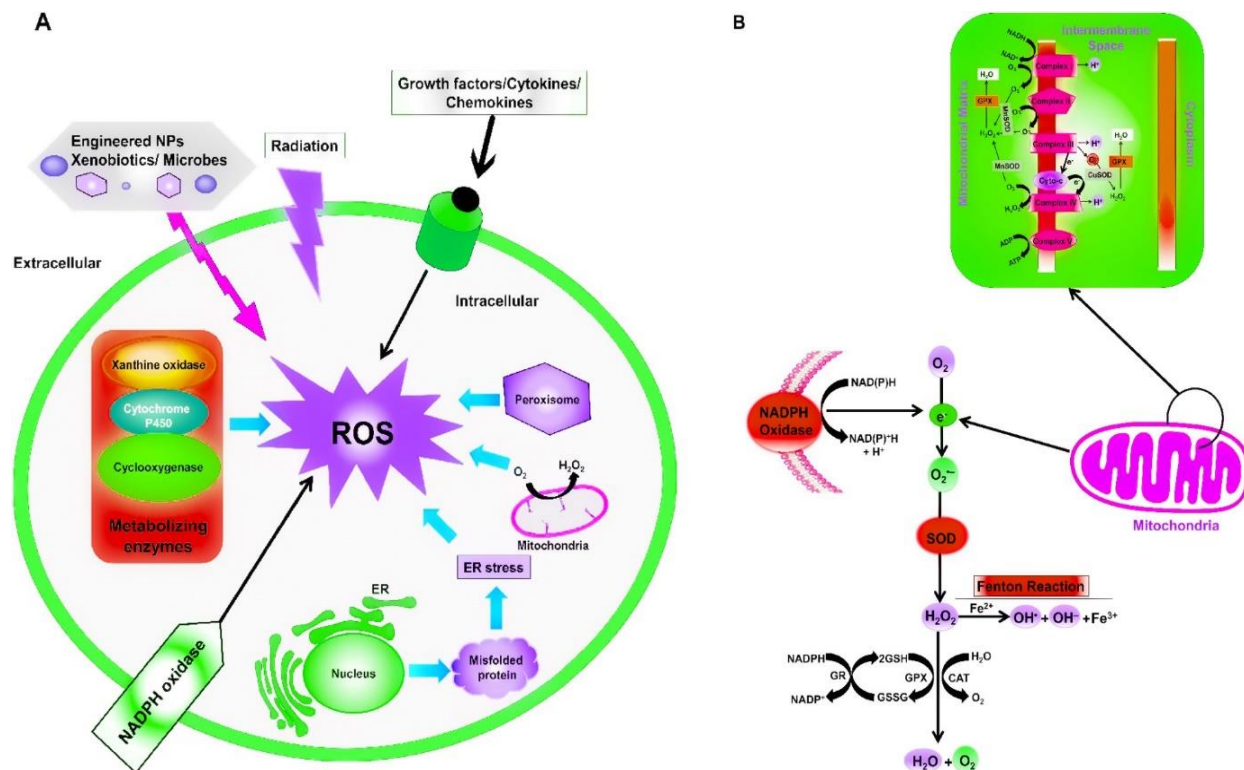


Figure 3. Generation sources of ROS. (a) A detailed diagram illustrating the origins of the generation of extracellular and intracellular ROS. Environmental contaminants, radiation exposure, microbial infections, and engineered nanoparticles' exposure are the extracellular causes of ROS (NPs). (b) A schematic diagram summarizes ROS formation from mitochondria, the mechanisms involved in ROS scavenging, and NADPH oxidase. To generate intracellular ROS, the NOX family, cell metabolizing enzymes, endoplasmic reticulum (ER) stress, and mitochondria can be generated. GPX: glutathione peroxidase; GSSG: Glutathione disulphide; e^- : electron; GR: glutathione reductase; Cyto-c: cytochrome-c; NOX:: CAT: catalase; SOD: superoxide dismutase and and NADPH oxidase [29].

$O_2^{\cdot-}$. Former research has shown that the reactive level of NPs impacts the formation of ROS [142]. The presence of free radicals and oxidants on the material surface influences ROS production. For instance, the ROS generation ($O_2^{\cdot-}$ and HO^{\cdot}) from quartz material is attributed to the attendance of Surface bounded radicals, like SiO_2^{\cdot} and SiO^{\cdot} [72, 116]. The capacity to control high amounts of oxidative stress is frequently improved with the Adsorption of suspended particles, such as nitrogen oxides and ozone, at the NP level [143].

Aqueous suspensions of quartz maters generate 1O_2 , HO^{\cdot} and H_2O_2 [75, 116, 142]. Metals and chemical compounds located at the NP surface have surface-dependent properties that increase the rate of ROS reactivity [136]. Different metal transitions, including vanadium, Cr, Cu, Si, and Fe, are involved in ROS production via the Fenton and Haber-Weiss reaction procedure [72]. The metal ion is oxidized, and HO is formed from the reaction of H_2O_2 with the metal ion in the Fenton reactions. H_2O_2 reduction using ferrous iron (Fe^{2+}) is also possible. Biological substances are also susceptible to the formation of HO and can be hazardous [76]. Metallic NPs, for instance, Fe and Cu, affect oxidative stress ($O_2^{\cdot-}$ and HO^{\cdot}) through Fenton reactions [87]; the reactions between oxidized metal ions and H_2O_2 to generate HO^{\cdot} are known as Haber-Weiss reactions [76, 144, 145]. NPs comprising Va, Cr, and Co equally Haber-Weiss and Fenton responses can be catalyzed [87], IONP-induced ROS generation is involved in Fenton and Fenton reactions [146]. A

unique feature of QDs-NPs, quantum limitation properties often play a critical role. Nanomaterials with quantum-confinement results have magnetic moments missing from the initial bulks [147]. Quantum confinement correspondingly changes NP affinity, causing them to donate or accept electrical charges, influencing their catalytic activities [147]. Atoms found on the NP level have lesser adjoining atoms than their original bulk content, theoretically reducing each atom's binding energy [147]. The Gibbs-Thomson equation predicts that the melting temperature of NP is lesser than that of bulk material [147]. Electrons are produced by various NPs activated by photon energy, including visible or ultraviolet radiation. [56, 148, 149]. This preserves the energy required to convert O_2 to 1O_2 , linked to cell damage via interactions through cellular lipids, proteins, and nucleic acids. [150].

7 ROS Production Induced by NP and Associated Cells

The cell membrane must be associated with NPs with specific characteristics that control the interchange of chemicals and ions from the outside world into the cell (Figure 3B). Endocytosis (cell transport) and exocytosis (cell transport) may transfer protein aggregates, lipoproteins, lipids, and nanomaterials to and from cells by encapsulating them within vesicles. Endocytosis can occur with or without clathrin or

caveolin protein, and it is critical for the internalization of NPs in the cell [151, 152]. Factors in the generation of NP-induced intracellular ROS contain free-radical reaction catalysis, interaction with mitochondrial components, growth factor activation, and NOX activation [152]. Mitochondria is a core organelle that is involved in the production of cellular ROS connected to NP. Interaction with the electron transfer chain and depolarization of the mitochondrial membrane are proven abilities of NPs [153, 154]. The mitochondrial electron transport chain is blocked after exposure to NPs, and the O₂ cell level rises due to electron transfer from the respiratory carriers to O₂. Human fibroblast cells and human glioblastoma exposed to AgNPs accumulated more AgNPs in mitochondria, disrupting the mitochondrial electron transport chain and causing substantial ROS-mediated cytotoxicity [154]. In *Escherichia coli*, the interaction of the dehydrogenase NADH by Ag ions was investigated, which inhibits the transport of electrons to O₂ and the production of numerous amounts of ROS [155].

Furthermore, NP exposure activates immune cells through a ROS-dependent pathway mediated by NOX [130]. The production of free radicals generated by NP aids in reducing glutathione disulfide, an oxidized form of GSH involved in oxidative stress and its repercussions [156, 157]. The activation of receptors and ROS-associated enzymes by NPs also contributes to the production of intracellular ROS caused by NPs. Metal oxide nanoparticles, for example (Ni₂O₃, Mn₂O₃, Co₃O₄, CoO, and Cr₂O₃ NPs) NADPH oxidation into cytochrome c and NADP⁺ oxidation, resulting in a significant amount of oxidative-stress-mediated poisonous [158]. The amounts of Energy required for band gaps associated with NPs are connected to this impact.

8 NP Production Induced by NP modulates biological Functions

Oxidative stress and the level of ROS produced, and that results are connected to the NMs concentrations that cells are exposed to [51]. Cells treated to minimal NP density demonstrated strong antioxidant advocacy proficient in withstanding oxidative damage and restoring redox equilibrium. On the other hand, high levels of NP overwhelm antioxidant mechanisms, leading to cytotoxicity and inflammation. Physiological activities such as photosynthesis, respiration, cell signaling, and ROS elements, for example, O₂^{•-}, HO[•], and H₂O₂, are critical mediates. Enzymes like CAT, SOD, and GPX and antioxidants like ascorbic acid, cysteine, and glutathione bilirubin all have a role in controlling their concentration within cells [159]. Various diseases can affect redox equilibrium, with oxidative stress resulting in cell damage owing to oxidative damage [160]. Changes in cell motility, uncontrolled cell signaling, cytotoxicity, apoptosis, DNA damage, and cancer growth and metastasis are all critical factors in nanotoxicity, as is oxidative stress [51-53]. ROS involvement in cell biological processes produced by NP and the molecular pathways involved are discussed in the subsections below (Figure 3B).

9 Stem Cell Biology Utilizing NP-induced Modulation in ROS Generation

Studies on the effect of metal NPs on MSC toxicity have been performed. Former research showed poisonous activity in bone marrow MSCs of ZnO NPs (BM-MSCs)[161]. The inconsistency of ZnONPs and the consequent secretion of Zn⁺ were caused by the acidic compartment containing lysosomal enzymes.

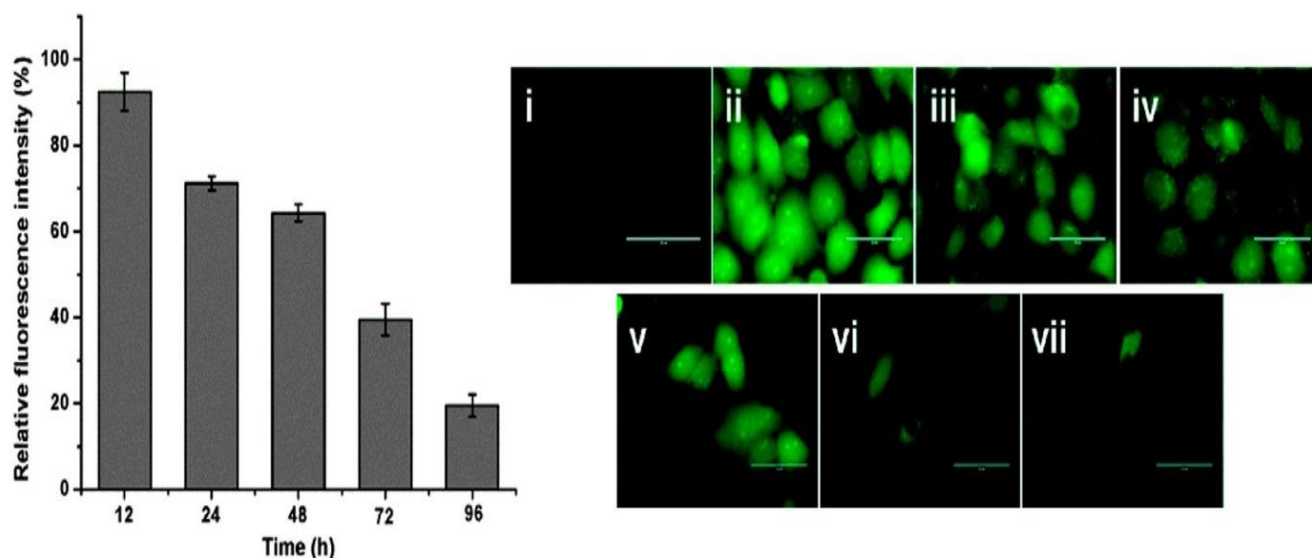


Figure 4: ROS controlling likely of nanoceria in H₂O₂ that exposure to L132. **I:** Uncontrolled cell type; **II:** Cells exposure to H₂O₂; **III** and **VII:** Pretreatment of time-dependent BCNPs in cells exposure to H₂O₂ [29, 167].

Furthermore, cytotoxicity in BM-MSCs associated with ZnONPs had a dose-dependent effect and was associated with ROS production. BM-MSC treatment using ZnONP, changes in apoptosis, and caspase-7/7 activation were also detected [161]. Results recently showed the ability of AgNPs in a ROS-dependent system to advance adipogenic separation of hMSCs (human MSCs) [162]. Increased expression of transcription factors specific to lipid differentiation and the presence of fat particles support AgNP-induced lipid differentiation. Furthermore, AgNPs' strong antibacterial activity enhanced hMSC adherence. Surprisingly, AgNPs had no negative effect on osteogenic hMSC development [162]. AgNPs may be a valuable nanomaterial for regulating ROS-related pathways in stem cell development and their potential application in tissue engineering and stem cell treatment. Inorganic antioxidants, nanoceria (cerium oxide NPs), can restrain free radicals with SOD and CAT-like processes [163-165]. Ce^{3+} and Ce^{4+} types are reduced at the nanoceria level located in the modulation of oxidation states [166]. In the human lung epithelial cell line (L-132) exposed to hydrogen peroxide (H_2O_2), nanoceria delivery through its encapsulation within the biological decomposable Alb-NPs to eventually create nanoceria encapsulated Alb-NPs (BCNPs) significantly scavenged ROS generation and thus avoided apoptotic changes (Figure 4) [167]. Nanoceria's antioxidant properties could also treat and regulate diseases caused by ROS's excessive development.

Nanoceria suppresses adipogenic growth in rat MSCs by reducing ROS production, which is required to differentiate fat cells from MSC. These NPs may be beneficial as NMs in the treatment of obesity [168]. Due to their ability to increase oxidative stress and disrupt redox equilibrium, recent research has documented the toxic activity of SPIONs in NSCs. Mitochondrial hyperpolarization, a significant reduction in the cell membrane breakage, intracellular GSH level, and increases in GPX and SOD levels have been shown by SPION-treated NSCs [169]. The effects of AgNPs coated with polysaccharides and hydrocarbons were investigated in another investigation. The proliferation and self-renewal of embryonic stem cells (ESCs) mediated by AgNP-driven ROS production were reduced by both chemically modified AgNPs. Even with cell cycle display at G1 / S stage by ESC-treated AgNPs, polysaccharide-coated AgNPs produce less ROS and had fewer negative effects than hydrocarbon-coated AgNPs [29, 170].

10 Conclusions

There has been an exponential increase in the development of engineered nanomaterials for industrial use, so the public has become interested in nanomaterials' safety and toxicity. Explaining a comprehensive biochemical mechanism of NMs-induced ROS production causing toxic effects is complicated due to free-radical creation and reactivity. The first line of defense against a hazard is the comprehension of the mechanism of NMs-induced toxicity. Various physical actuators may activate these MNPs to produce reactive oxygen species (ROS) that can be used to destroy bacteria and cancer cells selectively. For IONPs under AgNPs, and a magnetic field, AuNPs, TiO_2 NPs, and ZnONPs US exposure or under light, their central function as remotely activated NPs for the generation of remedial ROS has been emphasized. Consequently, outlook investigation should consist of MNPs focusing mainly on speeding up their therapeutic translation for biomedical use in systematic pre-clinical studies.

Ethical issue

Authors are aware of and comply with, best practices in publication ethics, specifically about authorship (avoidance of guest authorship), dual submission, manipulation of figures, competing interests, and compliance with policies on research ethics. Authors adhere to publication requirements that submitted work is original and has not been published elsewhere in any language.

Competing interests

The authors declare that no conflict of interest would prejudice the impartiality of this scientific work.

Authors contribution

All authors of this study have a complete contribution for data collection, data analyses, and manuscript writing.

References

1. Bayda, S., et al., *The history of nanoscience and nanotechnology: From chemical-physical applications to nanomedicine*. *Molecules*, 2020. **25**(1): p. 112.
2. Jeevanandam, J., et al., *Review on nanoparticles and nanostructured materials: history, sources, toxicity and regulations*. *Beilstein journal of nanotechnology*, 2018. **9**(1): p. 1050-1074.
3. Mazraeadoost, S. and G. Behbudi, *Basic Nano Magnetic Particles and Essential Oils: Biological Applications*. *Journal of Environmental Treatment Techniques*, 2021. **9**(3): p. 609-620.
4. Mazraeadoost, S. and G. Behbudi, *Nano materials-based devices by photodynamic therapy for treating cancer applications*. *Advances in Applied NanoBio-Technologies*, 2021. **2**(3): p. 9-21.
5. Masoumzade, R., G. Behbudi, and S. Mazraeadoost, *A medical encyclopedia with new approach graphene quantum dots for anti-breast cancer applications: mini review*. *Advances in Applied NanoBio-Technologies*, 2020. **1**(4): p. 84-90.
6. Mousavi, S.M., et al., *Data on cytotoxic and antibacterial activity of synthesized Fe₃O₄ nanoparticles using Malva sylvestris*. *Data in brief*, 2020. **28**: p. 104929.
7. Goudarzian, N., et al., *Enhancing the Physical, Mechanical, Oxygen Permeability and Photodegradation Properties of Styrene-acrylonitrile (SAN), Butadiene Rubber (BR) Composite by Silica Nanoparticles*. *Journal of Environmental Treatment Techniques*, 2020. **8**(2): p. 718-726.
8. Tech, J.E.T., *Investigating the Activity of Antioxidants Activities Content in Apiaceae and to Study Antimicrobial and Insecticidal Activity of Antioxidant by using SPME Fiber Assembly Carboxen/Polydimethylsiloxane (CAR/PDMS)*. *Journal of Environmental Treatment Techniques*, 2020. **8**(1): p. 214-24.
9. Bruchez, M., et al., *Semiconductor nanocrystals as fluorescent biological labels*. *science*, 1998. **281**(5385): p. 2013-2016.
10. Wang, S., et al., *Antigen/antibody immunocomplex from CdTe nanoparticle bioconjugates*. *Nano letters*, 2002. **2**(8): p. 817-822.
11. Ma, J., et al., *Biomimetic processing of nanocrystallite bioactive apatite coating on titanium*. *Nanotechnology*, 2003. **14**(6): p. 619.
12. Nam, J.-M., C.S. Thaxton, and C.A. Mirkin, *Nanoparticle-based bar codes for the ultrasensitive detection of proteins*. *science*, 2003. **301**(5641): p. 1884-1886.
13. Edelstein, R., et al., *The BARC biosensor applied to the detection of biological warfare agents*. *Biosensors and Bioelectronics*, 2000. **14**(10-

- 11): p. 805-813.
14. Mahtab, R., J.P. Rogers, and C.J. Murphy, *Protein-sized quantum dot luminescence can distinguish between "straight", "bent", and "kinked" oligonucleotides*. Journal of the American Chemical Society, 1995. **117**(35): p. 9099-9100.
15. Molday, R.S. and D. Mackenzie, *Immunospecific ferromagnetic iron-dextran reagents for the labeling and magnetic separation of cells*. Journal of immunological methods, 1982. **52**(3): p. 353-367.
16. Shen, Z., A. Wu, and X. Chen, *Iron oxide nanoparticle based contrast agents for magnetic resonance imaging*. Molecular pharmaceutics, 2017. **14**(5): p. 1352-1364.
17. Gordon, R., *Encyclopedia of Biophysics*. 2013.
18. Jin, S. and K. Ye, *Nanoparticle-mediated drug delivery and gene therapy*. Biotechnology progress, 2007. **23**(1): p. 32-41.
19. Gholami, A., et al., *3D nanostructures for tissue engineering, cancer therapy, and gene delivery*. Journal of Nanomaterials, 2020. **2020**.
20. Jaque, D., et al., *Nanoparticles for photothermal therapies*. nanoscale, 2014. **6**(16): p. 9494-9530.
21. Zhang, Y., et al., *Enhancement of HIFU ablation by sonosensitizer-loading liquid fluorocarbon nanoparticles with pre-targeting in a mouse model*. Scientific reports, 2019. **9**(1): p. 1-18.
22. Bosca, F., et al., *Exploiting Lipid and Polymer Nanocarriers to Improve the Anticancer Sonodynamic Activity of Chlorophyll*. Pharmaceutics, 2020. **12**(7): p. 605.
23. Brazzale, C., et al., *Enhanced selective sonosensitizing efficacy of ultrasound-based anticancer treatment by targeted gold nanoparticles*. Nanomedicine, 2016. **12**(23): p. 3053-3070.
24. Foglietta, F., et al., *Sonodynamic treatment as an innovative bimodal anticancer approach: shock wave-mediated tumor growth inhibition in a syngeneic breast cancer model*. 2015.
25. Varchi, G., et al., *Engineered porphyrin loaded core-shell nanoparticles for selective sonodynamic anticancer treatment*. Nanomedicine, 2015. **10**(23): p. 3483-3494.
26. Abrahamse, H., et al., *Nanoparticles for advanced photodynamic therapy of cancer*. Photomedicine and laser surgery, 2017. **35**(11): p. 581-588.
27. Abrahamse, H. and M.R. Hamblin, *New photosensitizers for photodynamic therapy*. Biochemical Journal, 2016. **473**(4): p. 347-364.
28. Sortino, S., *Light-responsive nanostructured systems for applications in nanomedicine*. 2016: Springer.
29. Abdal Dayem, A., et al., *The role of reactive oxygen species (ROS) in the biological activities of metallic nanoparticles*. International journal of molecular sciences, 2017. **18**(1): p. 120.
30. Ciccarese, F., et al., *Nanoparticles as tools to target redox homeostasis in cancer cells*. Antioxidants, 2020. **9**(3): p. 211.
31. Canaparo, R., et al., *Biomedical Applications of Reactive Oxygen Species Generation by Metal Nanoparticles*. Materials, 2021. **14**(1): p. 53.
32. Mousavi, S.M., et al., *Development of graphene based nanocomposites towards medical and biological applications*. Artificial cells, nanomedicine, and biotechnology, 2020. **48**(1): p. 1189-1205.
33. Mousavi, S.M., et al., *Nanosensors for chemical and biological and medical applications*. Med Chem (Los Angeles), 2018. **8**(8): p. 2161-0444.1000515.
34. Hashemi, S.A., et al., *Coupled graphene oxide with hybrid metallic nanoparticles as potential electrochemical biosensors for precise detection of ascorbic acid within blood*. Analytica chimica acta, 2020. **1107**: p. 183-192.
35. Bahrani, S., et al., *Zinc-based metal-organic frameworks as nontoxic and biodegradable platforms for biomedical applications: review study*. Drug metabolism reviews, 2019. **51**(3): p. 356-377.
36. Mousavi, M., et al., *Erythrosine adsorption from aqueous solution via decorated graphene oxide with magnetic iron oxide nano particles: kinetic and equilibrium studies*. Acta Chimica Slovenica, 2018. **65**(4): p. 882-894.
37. Avval, Z.M., et al., *Introduction of magnetic and supermagnetic nanoparticles in new approach of targeting drug delivery and cancer therapy application*. Drug metabolism reviews, 2020. **52**(1): p. 157-184.
38. Mousavi, S.M., et al., *Gold nanostars-diagnosis, bioimaging and biomedical applications*. Drug metabolism reviews, 2020. **52**(2): p. 299-318.
39. Mousavi, S.M., et al., *Carbon Substrates for Flexible Supercapacitors and Energy Storage Applications*. Flexible Supercapacitor Nanoarchitectonics, 2021: p. 95-141.
40. Raeisi, F., et al., *Application of biosurfactant as a demulsifying and emulsifying agent in the formulation of petrochemical products, in Green Sustainable Process for Chemical and Environmental Engineering and Science*. 2021, Elsevier. p. 399-422.
41. Mousavi, S.M., et al., *Development of hydrophobic reduced graphene oxide as a new efficient approach for photochemotherapy*. RSC Advances, 2020. **10**(22): p. 12851-12863.
42. Mousavi, S.M., et al., *Recent Advancements in Polythiophene-Based Materials and their Biomedical, Geno Sensor and DNA Detection*. International Journal of Molecular Sciences, 2021. **22**(13): p. 6850.
43. Mousavi, S.M., et al., *Recent Progress in Electrochemical Detection of Human Papillomavirus (HPV) via Graphene-Based Nanosensors*. Journal of Sensors, 2021. **2021**.
44. Mousavi, S.M., et al., *Asymmetric membranes: a potential scaffold for wound healing applications*. Symmetry, 2020. **12**(7): p. 1100.
45. Hashemi, S.A., et al., *Picomolar-level detection of mercury within non-biological/biological aqueous media using ultra-sensitive polyaniline-Fe₃O₄-silver diethyldithiocarbamate nanostructure*. Analytical and Bioanalytical Chemistry, 2020. **412**: p. 5353-5365.
46. Hashemi, S.A., et al., *Polythiophene silver bromide nanostructure as ultra-sensitive non-enzymatic electrochemical glucose biosensor*. European Polymer Journal, 2020. **138**: p. 109959.
47. Ahmadi, S., et al., *Anti-bacterial/fungal and anti-cancer performance of green synthesized Ag nanoparticles using summer savory extract*. Journal of Experimental Nanoscience, 2020. **15**(1): p. 363-380.
48. Hashemi, S.A., et al., *Integrated polyaniline with graphene oxide-iron tungsten nitride nanoflakes as ultrasensitive electrochemical sensor for precise detection of 4-nitrophenol within aquatic media*. Journal of Electroanalytical Chemistry, 2020. **873**: p. 114406.
49. Hashemi, S.A., et al., *Superior X-ray radiation shielding effectiveness of biocompatible polyaniline reinforced with hybrid graphene oxide-iron tungsten nitride flakes*. Polymers, 2020. **12**(6): p. 1407.
50. Mousavi, S.M., et al., *Green synthesis of supermagnetic Fe₃O₄-MgO nanoparticles via Nutmeg essential oil toward superior anti-bacterial and anti-fungal performance*. Journal of Drug Delivery Science and Technology, 2019. **54**: p. 101352.

51. Nel, A., et al., *Toxic potential of materials at the nanolevel*. science, 2006. **311**(5761): p. 622-627.
52. Xia, T., et al., *Comparison of the mechanism of toxicity of zinc oxide and cerium oxide nanoparticles based on dissolution and oxidative stress properties*. ACS nano, 2008. **2**(10): p. 2121-2134.
53. Zhu, X., et al., *Biosensing approaches for rapid genotoxicity and cytotoxicity assays upon nanomaterial exposure*. Small, 2013. **9**(9-10): p. 1821-1830.
54. Gonzalez, L., D. Lison, and M. Kirsch-Volders, *Genotoxicity of engineered nanomaterials: A critical review*. Nanotoxicology, 2008. **2**(4): p. 252-273.
55. Oberdörster, G., E. Oberdörster, and J. Oberdörster, *Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles*. Environ Health Perspect **113**: 823–839. 2005.
56. Li, Y., et al., *Chronic Al₂O₃-nanoparticle exposure causes neurotoxic effects on locomotion behaviors by inducing severe ROS production and disruption of ROS defense mechanisms in nematode *Caenorhabditis elegans**. Journal of hazardous materials, 2012. **219**: p. 221-230.
57. Winnik, F.M. and D. Maysinger, *Quantum dot cytotoxicity and ways to reduce it*. Accounts of chemical research, 2013. **46**(3): p. 672-680.
58. Akhtar, M.J., et al., *Nanotoxicity of pure silica mediated through oxidant generation rather than glutathione depletion in human lung epithelial cells*. Toxicology, 2010. **276**(2): p. 95-102.
59. Akhtar, M.J., et al., *Protective effect of sulphoraphane against oxidative stress mediated toxicity induced by CuO nanoparticles in mouse embryonic fibroblasts BALB 3T3*. The Journal of toxicological sciences, 2012. **37**(1): p. 139-148.
60. Fan, Z. and J.G. Lu, *Zinc oxide nanostructures: synthesis and properties*. Journal of nanoscience and nanotechnology, 2005. **5**(10): p. 1561-1573.
61. Chiang, H.-m., et al., *Nanoscale ZnO induces cytotoxicity and DNA damage in human cell lines and rat primary neuronal cells*. Journal of nanoscience and nanotechnology, 2012. **12**(3): p. 2126-2135.
62. Yin, J.-J., et al., *Electron spin resonance spectroscopy for studying the generation and scavenging of reactive oxygen species by nanomaterials*, in *Nanopharmaceutics: The Potential Application of Nanomaterials*. 2013, World Scientific. p. 375-400.
63. Yin, J.-J., et al., *Phototoxicity of nano titanium dioxides in HaCaT keratinocytes—generation of reactive oxygen species and cell damage*. Toxicology and applied pharmacology, 2012. **263**(1): p. 81-88.
64. Applerot, G., et al., *Enhanced antibacterial activity of nanocrystalline ZnO due to increased ROS-mediated cell injury*. Advanced Functional Materials, 2009. **19**(6): p. 842-852.
65. Wang, J.J., B.J. Sanderson, and H. Wang, *Cyto-and genotoxicity of ultrafine TiO₂ particles in cultured human lymphoblastoid cells*. Mutation Research/Genetic Toxicology and Environmental Mutagenesis, 2007. **628**(2): p. 99-106.
66. Girgis, E., et al., *Nanotoxicity of gold and gold–cobalt nanoalloy*. Chemical research in toxicology, 2012. **25**(5): p. 1086-1098.
67. Hsin, Y.-H., et al., *The apoptotic effect of nanosilver is mediated by a ROS-and JNK-dependent mechanism involving the mitochondrial pathway in NIH3T3 cells*. Toxicology letters, 2008. **179**(3): p. 130-139.
68. Kim, S. and D.Y. Ryu, *Silver nanoparticle-induced oxidative stress, genotoxicity and apoptosis in cultured cells and animal tissues*. Journal of applied toxicology, 2013. **33**(2): p. 78-89.
69. Shvedova, A., et al., *Exposure to carbon nanotube material: assessment of nanotube cytotoxicity using human keratinocyte cells*. Journal of toxicology and environmental health Part A, 2003. **66**(20): p. 1909-1926.
70. Liu, Y., et al., *Plastic protein microarray to investigate the molecular pathways of magnetic nanoparticle-induced nanotoxicity*. Nanotechnology, 2013. **24**(17): p. 175501.
71. Wang, Y., et al., *A study of the mechanism of in vitro cytotoxicity of metal oxide nanoparticles using catfish primary hepatocytes and human HepG2 cells*. Science of the total environment, 2011. **409**(22): p. 4753-4762.
72. Knaapen, A.M., et al., *Inhaled particles and lung cancer. Part A: Mechanisms*. International Journal of Cancer, 2004. **109**(6): p. 799-809.
73. Fu, P.P., et al., *Mechanisms of nanotoxicity: generation of reactive oxygen species*. Journal of food and drug analysis, 2014. **22**(1): p. 64-75.
74. Manke, A., L. Wang, and Y. Rojanasakul, *Mechanisms of nanoparticle-induced oxidative stress and toxicity*. BioMed research international, 2013. **2013**.
75. Vallyathan, V. and X. Shi, *The role of oxygen free radicals in occupational and environmental lung diseases*. Environmental Health Perspectives, 1997. **105**(suppl 1): p. 165-177.
76. Thannickal, V.J. and B.L. Fanburg, *Reactive oxygen species in cell signaling*. American Journal of Physiology-Lung Cellular and Molecular Physiology, 2000. **279**(6): p. L1005-L1028.
77. Maryanovich, M. and A. Gross, *A ROS rheostat for cell fate regulation*. Trends in cell biology, 2013. **23**(3): p. 129-134.
78. Sena, L.A. and N.S. Chandel, *Physiological roles of mitochondrial reactive oxygen species*. Molecular cell, 2012. **48**(2): p. 158-167.
79. Cremers, C.M. and U. Jakob, *Oxidant sensing by reversible disulfide bond formation*. Journal of Biological Chemistry, 2013. **288**(37): p. 26489-26496.
80. Snezhkina, A.V., et al., *ROS generation and antioxidant defense systems in normal and malignant cells*. Oxidative medicine and cellular longevity, 2019. **2019**.
81. Riley, P., *Free radicals in biology: oxidative stress and the effects of ionizing radiation*. International journal of radiation biology, 1994. **65**(1): p. 27-33.
82. Birben, E., et al., *Oxidative stress and antioxidant defense*. World Allergy Organization Journal, 2012. **5**(1): p. 9-19.
83. Poljsak, B., D. Šuput, and I. Milisav, *Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants*. Oxidative medicine and cellular longevity, 2013. **2013**.
84. Forrester, S.J., et al., *Reactive oxygen species in metabolic and inflammatory signaling*. Circulation research, 2018. **122**(6): p. 877-902.
85. Dunnyaporn, T., et al., *Redox regulation of cell survival*. Antioxid Redox Signal, 2008. **10**(8): p. p1343-1374.
86. Bae, Y.S., et al., *Regulation of reactive oxygen species generation in cell signaling*. Molecules and cells, 2011. **32**(6): p. 491-509.
87. Valko, M., et al., *Free radicals, metals and antioxidants in oxidative stress-induced cancer*. Chemo-biological interactions, 2006. **160**(1): p. 1-40.
88. Touyz, R.M., *Molecular and cellular mechanisms in vascular injury in hypertension: role of angiotensin II—editorial review*. Current opinion in nephrology and hypertension, 2005. **14**(2): p. 125-131.
89. Mueller, C., *Laude K, McNally JS, Harrison DG. ATVB in focus: redox mechanisms in blood vessels*. Arterioscler Thromb Vasc Biol,

2005. **25**: p. 274-278.
90. Wu, H., et al., *Reactive oxygen species-related activities of nano-iron metal and nano-iron oxides*. Journal of Food and Drug Analysis, 2014. **22**(1): p. 86-94.
91. Halliwell, B., *Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life*. Plant physiology, 2006. **141**(2): p. 312-322.
92. Johnson, F. and C. Giulivi, *Superoxide dismutases and their impact upon human health*. Molecular aspects of medicine, 2005. **26**(4-5): p. 340-352.
93. Paravicini, T.M. and R.M. Touyz, *NADPH oxidases, reactive oxygen species, and hypertension: clinical implications and therapeutic possibilities*. Diabetes care, 2008. **31**(Supplement 2): p. S170-S180.
94. Brand, M.D., *The sites and topology of mitochondrial superoxide production*. Experimental gerontology, 2010. **45**(7-8): p. 466-472.
95. Halliwell, B., M.V. Clement, and L.H. Long, *Hydrogen peroxide in the human body*. FEBS letters, 2000. **486**(1): p. 10-13.
96. Hampton, M.B. and S. Orrenius, *Dual regulation of caspase activity by hydrogen peroxide: implications for apoptosis*. FEBS letters, 1997. **414**(3): p. 552-556.
97. Ray, P.D., B.-W. Huang, and Y. Tsuji, *Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling*. Cellular signalling, 2012. **24**(5): p. 981-990.
98. Guzik, T.J. and D.G. Harrison, *Vascular NADPH oxidases as drug targets for novel antioxidant strategies*. Drug discovery today, 2006. **11**(11-12): p. 524-533.
99. Coso, S., et al., *NADPH oxidases as regulators of tumor angiogenesis: current and emerging concepts*. Antioxidants & redox signaling, 2012. **16**(11): p. 1229-1247.
100. Bedard, K. and K.-H. Krause, *The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology*. Physiological reviews, 2007. **87**(1): p. 245-313.
101. Storz, P., *Forkhead homeobox type O transcription factors in the responses to oxidative stress*. Antioxidants & redox signaling, 2011. **14**(4): p. 593-605.
102. Salata, O.V., *Applications of nanoparticles in biology and medicine*. Journal of nanobiotechnology, 2004. **2**(1): p. 1-6.
103. Brar, S.K., et al., *Engineered nanoparticles in wastewater and wastewater sludge—Evidence and impacts*. Waste management, 2010. **30**(3): p. 504-520.
104. Ray, P.C., H. Yu, and P.P. Fu, *Nanogold-based sensing of environmental toxins: excitement and challenges*. Journal of Environmental Science and Health, Part C, 2011. **29**(1): p. 52-89.
105. Poljak-Blaži, M., M. Jaganjac, and N. Žarković, *Cell oxidative stress: risk of metal nanoparticles*. Handbook of Nanophysics Nanomedicine and Nanorobotics, 2010: p. 16-1-16-17.
106. Röcker, C., et al., *A quantitative fluorescence study of protein monolayer formation on colloidal nanoparticles*. Nature nanotechnology, 2009. **4**(9): p. 577-580.
107. Jiang, X., et al., *Quantitative analysis of the protein corona on FePt nanoparticles formed by transferrin binding*. Journal of The Royal Society Interface, 2010. **7**(suppl_1): p. S5-S13.
108. Lynch, I. and K.A. Dawson, *Protein-nanoparticle interactions*. Nano today, 2008. **3**(1-2): p. 40-47.
109. Li, N., T. Xia, and A.E. Nel, *The role of oxidative stress in ambient particulate matter-induced lung diseases and its implications in the toxicity of engineered nanoparticles*. Free radical biology and medicine, 2008. **44**(9): p. 1689-1699.
110. Stone, V., H. Johnston, and M.J. Clift, *Air pollution, ultrafine and nanoparticle toxicology: cellular and molecular interactions*. IEEE transactions on nanobioscience, 2007. **6**(4): p. 331-340.
111. Shvedova, A.A., et al., *Mechanisms of carbon nanotube-induced toxicity: focus on oxidative stress*. Toxicology and applied pharmacology, 2012. **261**(2): p. 121-133.
112. Zhang, Z., et al., *On the interactions of free radicals with gold nanoparticles*. Journal of the American Chemical Society, 2003. **125**(26): p. 7959-7963.
113. Kennedy, I.M., D. Wilson, and A.I. Barakat, *Uptake and inflammatory effects of nanoparticles in a human vascular endothelial cell line*. Research Report (Health Effects Institute), 2009(136): p. 3-32.
114. Devarajan, P.V. and S. Jain, *Targeted drug delivery: concepts and design*. 2015: Springer.
115. Huang, Y.-W., C.-h. Wu, and R.S. Aronstam, *Toxicity of transition metal oxide nanoparticles: recent insights from in vitro studies*. Materials, 2010. **3**(10): p. 4842-4859.
116. Fubini, B. and A. Hubbard, *Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis*. Free Radical Biology and Medicine, 2003. **34**(12): p. 1507-1516.
117. Trachootham, D., J. Alexandre, and P. Huang, *Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach?* Nature reviews Drug discovery, 2009. **8**(7): p. 579-591.
118. Finkel, T., *Signal transduction by mitochondrial oxidants*. Journal of Biological Chemistry, 2012. **287**(7): p. 4434-4440.
119. Dikalov, S., *Cross talk between mitochondria and NADPH oxidases*. Free Radical Biology and Medicine, 2011. **51**(7): p. 1289-1301.
120. Tahara, E.B., F.D. Navarete, and A.J. Kowaltowski, *Tissue-, substrate-, and site-specific characteristics of mitochondrial reactive oxygen species generation*. Free Radical Biology and Medicine, 2009. **46**(9): p. 1283-1297.
121. Okado-Matsumoto, A. and I. Fridovich, *Subcellular distribution of superoxide dismutases (SOD) in rat liver: Cu, Zn-SOD in mitochondria*. Journal of Biological Chemistry, 2001. **276**(42): p. 38388-38393.
122. Murphy, M.P., *How mitochondria produce reactive oxygen species*. Biochemical journal, 2009. **417**(1): p. 1-13.
123. Finkel, T., *Signal transduction by reactive oxygen species*. Journal of Cell Biology, 2011. **194**(1): p. 7-15.
124. MacFie, T.S., et al., *DUOX2 and DUOX2 form the predominant enzyme system capable of producing the reactive oxygen species H2O2 in active ulcerative colitis and are modulated by 5-aminosalicylic acid*. Inflammatory bowel diseases, 2014. **20**(3): p. 514-524.
125. Yoshihara, A., et al., *Regulation of dual oxidase expression and H2O2 production by thyroglobulin*. Thyroid, 2012. **22**(10): p. 1054-1062.
126. van der Vlies, D., et al., *Oxidation of ER resident proteins upon oxidative stress: effects of altering cellular redox/antioxidant status and implications for protein maturation*. Antioxidants and redox signaling, 2003. **5**(4): p. 381-387.
127. Yasmin, W., K.D. Strynadka, and R. Schulz, *Generation of peroxynitrite contributes to ischemia-reperfusion injury in isolated rat hearts*. Cardiovascular research, 1997. **33**(2): p. 422-432.

128. Halliwell, B. and J.M. Gutteridge, *Reactive species can be useful: some more examples*, in *Free Radicals in Biology and Medicine*. 2015, Oxford University Press.
129. Mignolet-Spruyt, L., et al., *Spreading the news: subcellular and organellar reactive oxygen species production and signalling*. *Journal of experimental botany*, 2016. **67**(13): p. 3831-3844.
130. Risom, L., P. Møller, and S. Loft, *Oxidative stress-induced DNA damage by particulate air pollution*. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 2005. **592**(1-2): p. 119-137.
131. Bonner, J.C., *Lung fibrotic responses to particle exposure*. *Toxicologic pathology*, 2007. **35**(1): p. 148-153.
132. Ray, P.C., H. Yu, and P.P. Fu, *Toxicity and environmental risks of nanomaterials: challenges and future needs*. *Journal of Environmental Science and Health Part C*, 2009. **27**(1): p. 1-35.
133. Wang, S., et al., *Challenge in understanding size and shape dependent toxicity of gold nanomaterials in human skin keratinocytes*. *Chemical physics letters*, 2008. **463**(1-3): p. 145-149.
134. Shaligram, S. and A. Campbell, *Toxicity of copper salts is dependent on solubility profile and cell type tested*. *Toxicology in Vitro*, 2013. **27**(2): p. 844-851.
135. Lu, W., et al., *Effect of surface coating on the toxicity of silver nanomaterials on human skin keratinocytes*. *Chemical physics letters*, 2010. **487**(1-3): p. 92-96.
136. Wilson, M.R., et al., *Interactions between ultrafine particles and transition metals in vivo and in vitro*. *Toxicology and applied pharmacology*, 2002. **184**(3): p. 172-179.
137. Sioutas, C., R.J. Delfino, and M. Singh, *Exposure assessment for atmospheric ultrafine particles (UFPs) and implications in epidemiologic research*. *Environmental health perspectives*, 2005. **113**(8): p. 947-955.
138. Stone, V., et al., *The role of oxidative stress in the prolonged inhibitory effect of ultrafine carbon black on epithelial cell function*. *Toxicology in vitro*, 1998. **12**(6): p. 649-659.
139. Fan, J., et al., *Direct evidence for catalase and peroxidase activities of ferritin-platinum nanoparticles*. *Biomaterials*, 2011. **32**(6): p. 1611-1618.
140. Donaldson, K. and C.L. Tran, *Inflammation caused by particles and fibers*. *Inhalation toxicology*, 2002. **14**(1): p. 5-27.
141. Oberdörster, G., et al., *Principles for characterizing the potential human health effects from exposure to nanomaterials: elements of a screening strategy*. *Particle and fibre toxicology*, 2005. **2**(1): p. 1-35.
142. Schins, R.P., *Mechanisms of genotoxicity of particles and fibers*. *Inhalation toxicology*, 2002. **14**(1): p. 57-78.
143. Buzea, C., I.I. Pacheco, and K. Robbie, *Nanomaterials and nanoparticles: sources and toxicity*. *Biointerphases*, 2007. **2**(4): p. MR17-MR71.
144. Aust, S., et al., *Free radicals in toxicology*. *Toxicology and applied pharmacology*, 1993. **120**(2): p. 168-178.
145. Mousavi, S.M., et al., *Precise Blood Glucose Sensing by Nitrogen-Doped Graphene Quantum Dots for Tight Control of Diabetes*. *Journal of Sensors*, 2021. **2021**.
146. Fang, G.-D., D.-M. Zhou, and D.D. Dionysiou, *Superoxide mediated production of hydroxyl radicals by magnetite nanoparticles: demonstration in the degradation of 2-chlorobiphenyl*. *Journal of Hazardous Materials*, 2013. **250**: p. 68-75.
147. Roduner, E., *Size matters: why nanomaterials are different*. *Chemical Society Reviews*, 2006. **35**(7): p. 583-592.
148. Yaghini, E., et al. *Reactive oxygen species generation from photoexcited quantum dot nanoparticles: Type I versus type II photochemical mechanism*. 2011. 13th International Photodynamic Association (IPA) World Congress.
149. Mousavi, S.M., et al., *Multifunctional Gold Nanorod for Therapeutic Applications and Pharmaceutical Delivery Considering Cellular Metabolic Responses, Oxidative Stress and Cellular Longevity*. *Nanomaterials*, 2021. **11**(7): p. 1868.
150. Sharma, P., et al., *Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions*. *Journal of botany*, 2012. **2012**.
151. Iversen, T.-G., T. Skotland, and K. Sandvig, *Endocytosis and intracellular transport of nanoparticles: Present knowledge and need for future studies*. *Nano today*, 2011. **6**(2): p. 176-185.
152. Canton, I. and G. Battaglia, *Endocytosis at the nanoscale*. *Chemical Society Reviews*, 2012. **41**(7): p. 2718-2739.
153. Smith, K.R., L.R. Klei, and A. Barchowsky, *Arsenite stimulates plasma membrane NADPH oxidase in vascular endothelial cells*. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 2001. **280**(3): p. L442-L449.
154. Xia, T., et al., *Comparison of the abilities of ambient and manufactured nanoparticles to induce cellular toxicity according to an oxidative stress paradigm*. *Nano letters*, 2006. **6**(8): p. 1794-1807.
155. AshaRani, P., *Low Kah Mun G, Hande MP, Valiyaveetil S*. Cytotoxicity and genotoxicity of silver nanoparticles in human cells. *ACS Nano*, 2009. **3**(2): p. 279-290.
156. Rahman, K., *Studies on free radicals, antioxidants, and co-factors*. *Clinical interventions in aging*, 2007. **2**(2): p. 219.
157. Fenoglio, I., et al., *The oxidation of glutathione by cobalt/tungsten carbide contributes to hard metal-induced oxidative stress*. *Free radical research*, 2008. **42**(8): p. 437-745.
158. Zhang, H., et al., *Use of metal oxide nanoparticle band gap to develop a predictive paradigm for oxidative stress and acute pulmonary inflammation*. *ACS nano*, 2012. **6**(5): p. 4349-4368.
159. Xu, Q., et al., *Reactive oxygen species (ROS) responsive polymers for biomedical applications*. *Macromolecular bioscience*, 2016. **16**(5): p. 635-646.
160. Tapeinos, C. and A. Pandit, *Physical, chemical, and biological structures based on ROS-sensitive moieties that are able to respond to oxidative microenvironments*. *Advanced Materials*, 2016. **28**(27): p. 5553-5585.
161. Syama, S., et al., *Zinc oxide nanoparticles induced oxidative stress in mouse bone marrow mesenchymal stem cells*. *Toxicology mechanisms and methods*, 2014. **24**(9): p. 644-653.
162. He, W., et al., *Silver nanoparticle based coatings enhance adipogenesis compared to osteogenesis in human mesenchymal stem cells through oxidative stress*. *Journal of Materials Chemistry B*, 2016. **4**(8): p. 1466-1479.
163. Celardo, I., et al., *Ce³⁺ ions determine redox-dependent anti-apoptotic effect of cerium oxide nanoparticles*. *ACS nano*, 2011. **5**(6): p. 4537-4549.
164. Heckert, E.G., et al., *The role of cerium redox state in the SOD mimetic activity of nanoceria*. *Biomaterials*, 2008. **29**(18): p. 2705-2709.
165. Celardo, I., E. Traversa, and L. Ghibelli, *Cerium oxide nanoparticles: a promise for applications in therapy*. *J Exp Ther Oncol*,

2011. **9**(1): p. 47-51.

166. Karakoti, A.S., et al., *Preparation and characterization challenges to understanding environmental and biological impacts of ceria nanoparticles*. Surface and Interface Analysis, 2012. **44**(8): p. 882-889.

167. Bhushan, B. and P. Gopinath, *Antioxidant nanozyme: a facile synthesis and evaluation of the reactive oxygen species scavenging potential of nanoceria encapsulated albumin nanoparticles*. Journal of Materials Chemistry B, 2015. **3**(24): p. 4843-4852.

168. Rocca, A., et al., *Cerium oxide nanoparticles inhibit adipogenesis in rat mesenchymal stem cells: potential therapeutic implications*. Pharmaceutical research, 2014. **31**(11): p. 2952-2962.

169. Pongrac, I.M., et al., *Oxidative stress response in neural stem cells exposed to different superparamagnetic iron oxide nanoparticles*. International journal of nanomedicine, 2016. **11**: p. 1701.

170. Rajanahalli, P., C. Stucke, and Y. Hong, *The effects of silver nanoparticles on mouse embryonic stem cell self-renewal and proliferation*, *Toxicol. Rep. 2* (2015) 758–764.